

Validation of a triple stage *in vitro* human gut model to study the biofilm mode of growth of *C. difficile* and the indigenous gut microbiota

Background

- ❑ Human gastrointestinal tract harbours complex microbial communities – **planktonic** and **biofilm**
- ❑ **Biofilm** - aggregation of bacteria bound to a substratum material and encased in a self produced extra-cellular matrix
- ❑ Bacteria within biofilms differ from planktonic bacteria:
- ❑ *C. difficile* adheres to colonic cells and mucus*
- ❑ Little else known about presence of CD in biofilms

*Gomez-Trevino, M *et al* 1996; Karjalainen, T *et al* 1994

In vitro human gut model

- ❑ Successfully validated and well established model in which to study CDI
- ❑ Successfully used to:
 - compare the efficacies of therapies
 - induce CDI
 - study effect of antimicrobial administration on the human gut microbiota

In vitro human gut model

- ❑ Complex growth medium
- ❑ Pooled human faecal emulsion
- ❑ Anaerobic
- ❑ 37 °C
- ❑ pH 5.5 – 6.8
- ❑ Planktonic indigenous gut microbiota and CD populations monitored



Proximal
colon



Distal
colon

In vitro human gut model

- ☐ Biofilm present on walls of vessel within the original gut model
- ☐ Spikes in planktonic bacterial populations
- ☐ Biofilm shedding?
- ☐ CD present?
- ☐ Treatment failure/recurrence?

In vitro human gut model

- Sampled biofilm at end of experimental run and compared to planktonic:
 - ▣ CD germination, proliferation, toxin production and treatment

	Toxin
Planktonic	✗
Biofilm	✓

- Cytotoxin persistence?
- CD toxin producing strain present within biofilm?

Aims

- ❑ Adapt human gut model to study planktonic and sessile bacterial populations
- ❑ Facilitate the formation of biofilm
- ❑ Easily sample sessile and planktonic populations longitudinally
- ❑ Evaluate role of biofilms in treatment failure/recurrence of CDI

Investigational/preliminary work

❑ **Biofilm scraping**

- Mature biofilm, difficult to sample

❑ **Mucin alginate beads**

- Realistic substratum material
- Degradation

❑ **Glass beads**

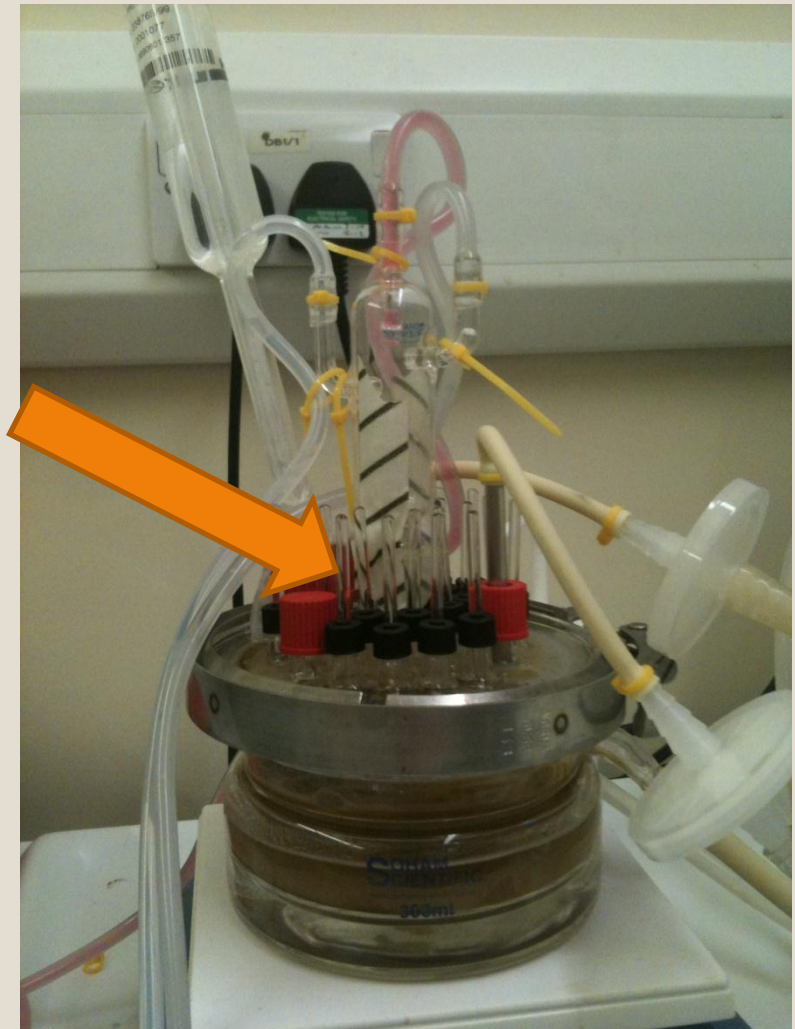
- Difficulty sampling

❑ **Glass rods**

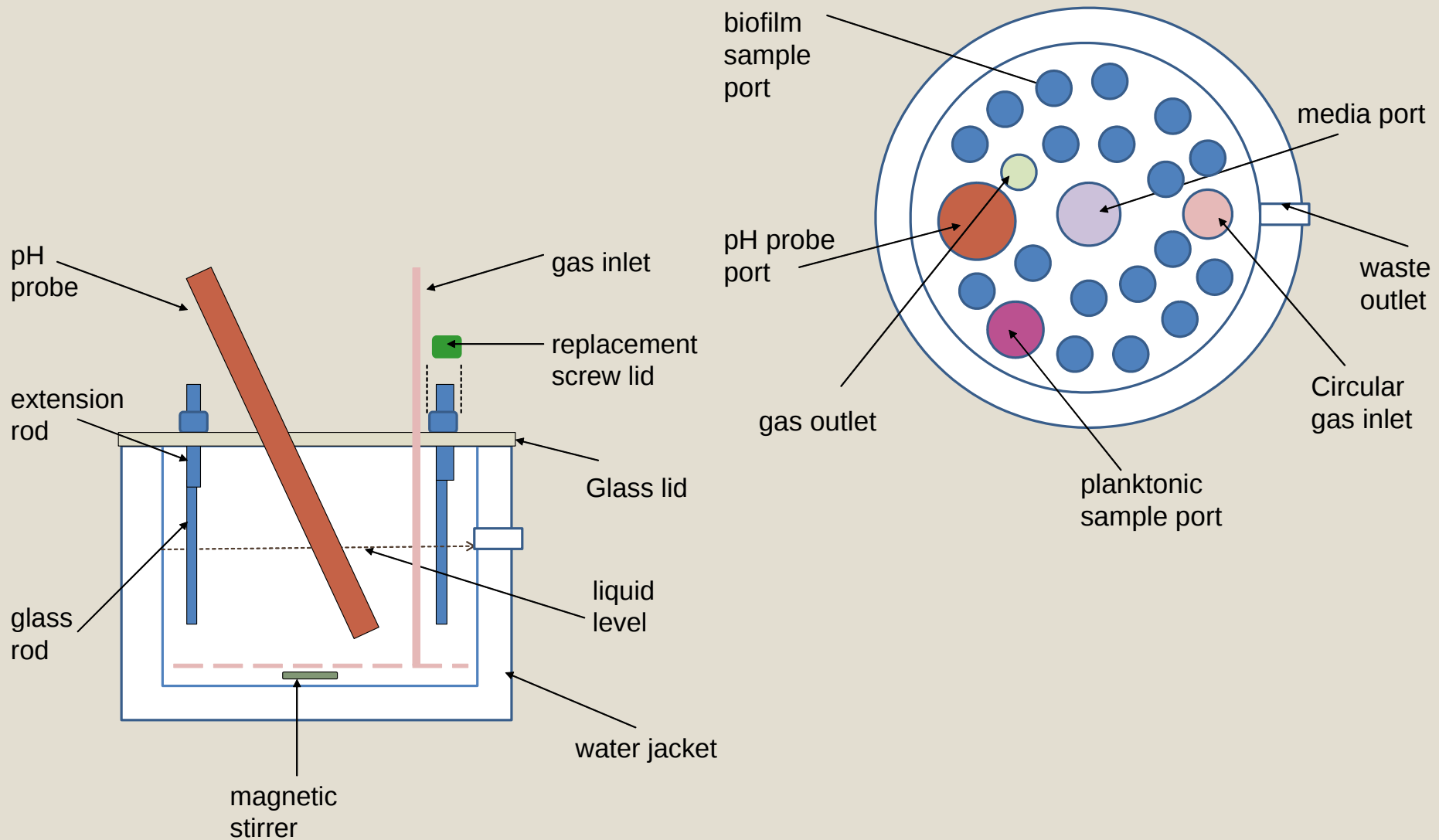
- Increased biofilm on liquid/air interface

Biofilm human gut model

- ❑ Adapt original gut model
- ❑ Maintain basic vessel design
- ❑ Maximise number of rods/sample points
- ❑ Ensure biofilm can be sampled with ease



Biofilm human gut model

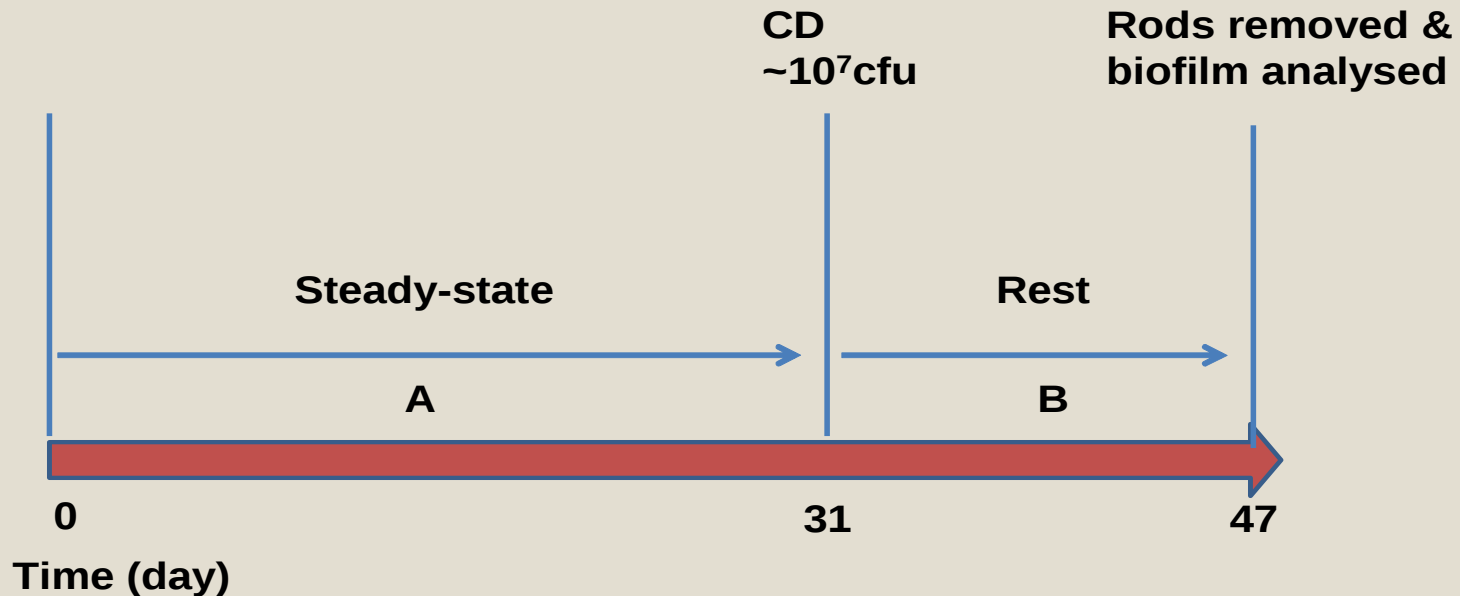


Biofilm human gut model

- ❑ Validation of a human gut model to facilitate and monitor the formation of biofilm
 - Ensure biofilm forms on all rods
 - Ensure consistency of biofilm populations across all rods
 - Ensure reproducibility of model
 - Ensure that planktonic populations behave as they did in original gut model

Biofilm human gut model - methods

- ❑ Single stage model
- ❑ 18 rods



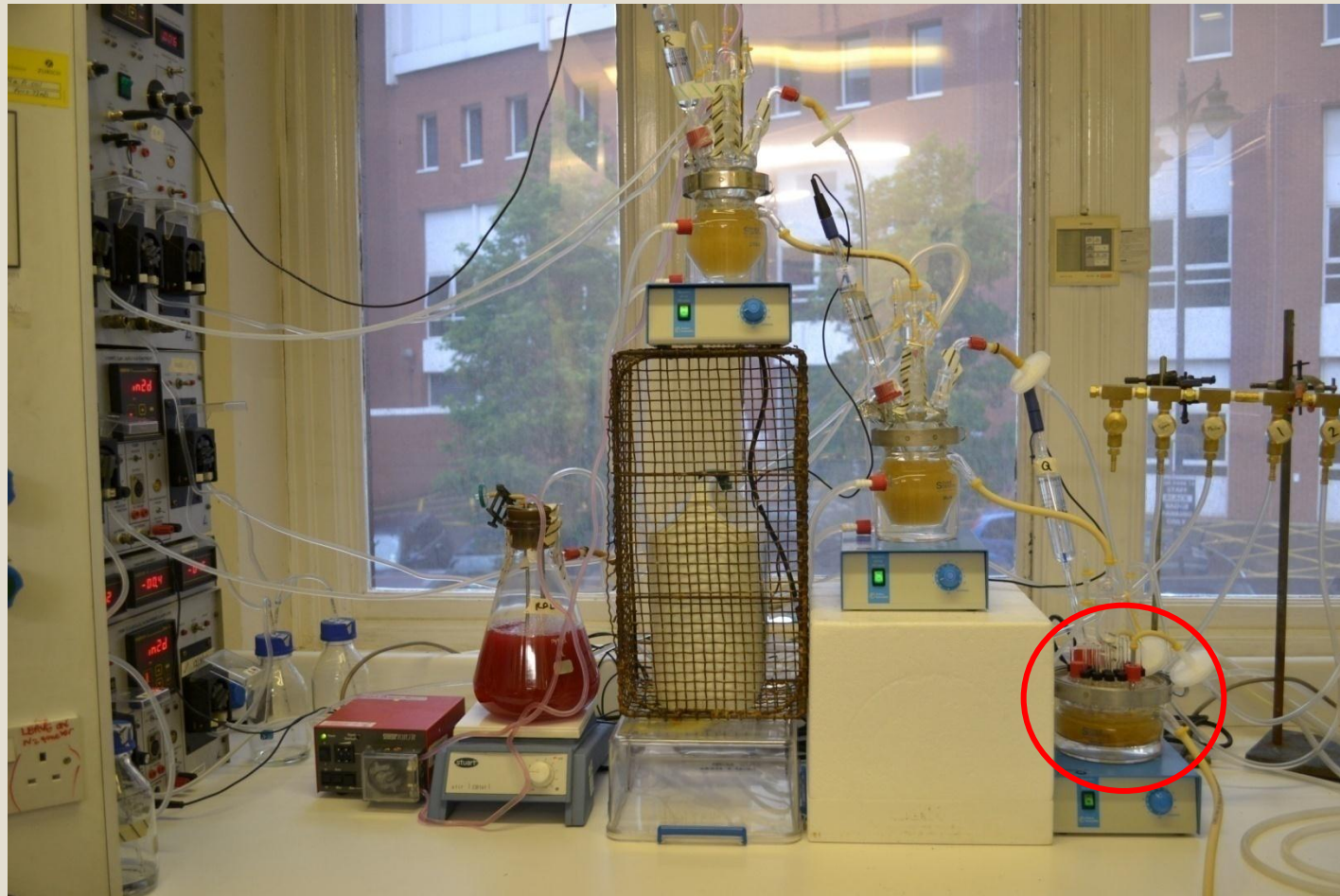
- ❑ Monitor planktonic populations throughout
- ❑ Experiment repeated on separate occasion

Biofilm human gut model - results

Results

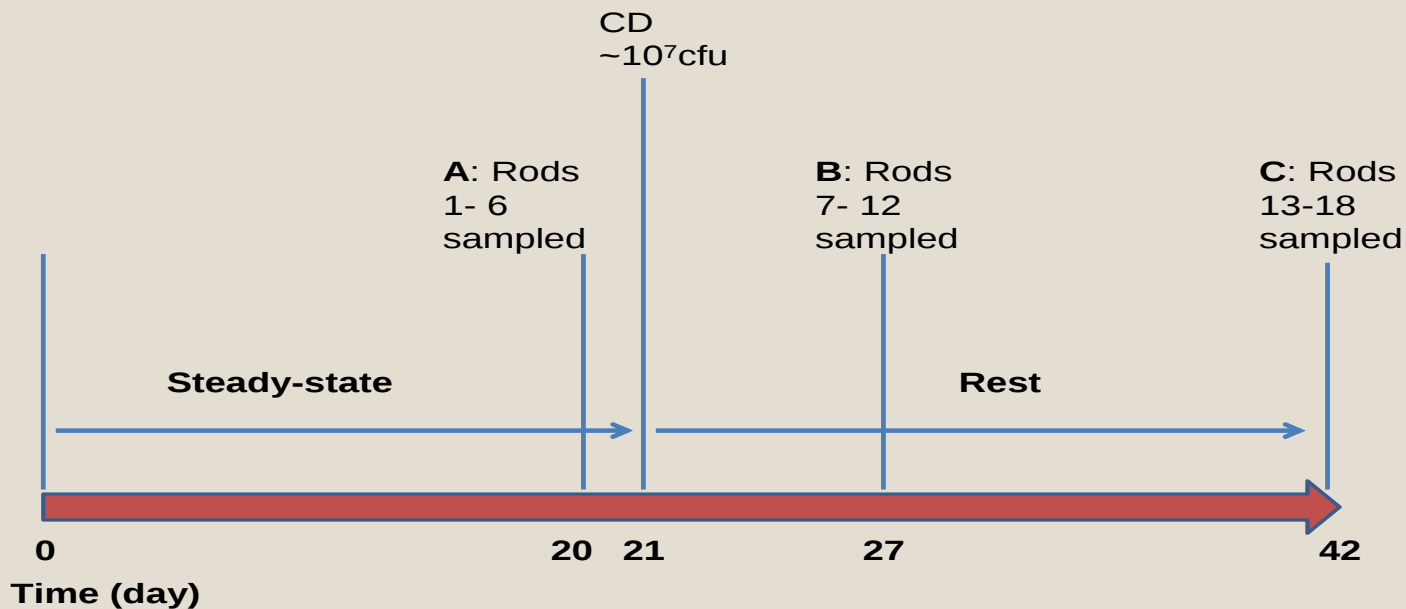
- ❑ All glass rods facilitated biofilm formation
- ❑ Planktonic and sessile populations were comparable including *C. difficile*
- ❑ Variation of total biofilm biomass amongst the rods
- ❑ Populations of each bacterial group in $\log_{10}\text{cfu/g}$ had minimal variation between rods
- ❑ Repeat experiment had similar results

Triple stage biofilm human gut model



Triple stage biofilm human gut model

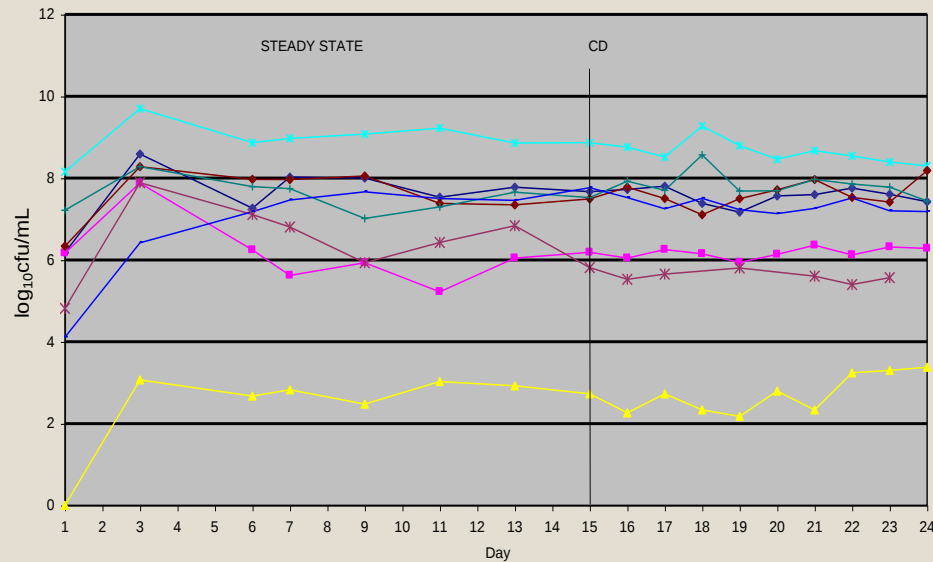
- ❑ Vessel design same as in single stage



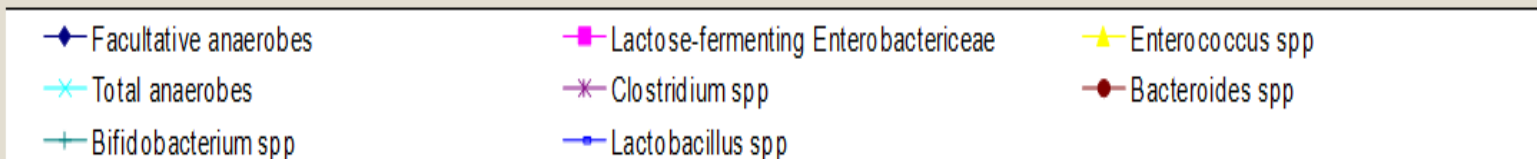
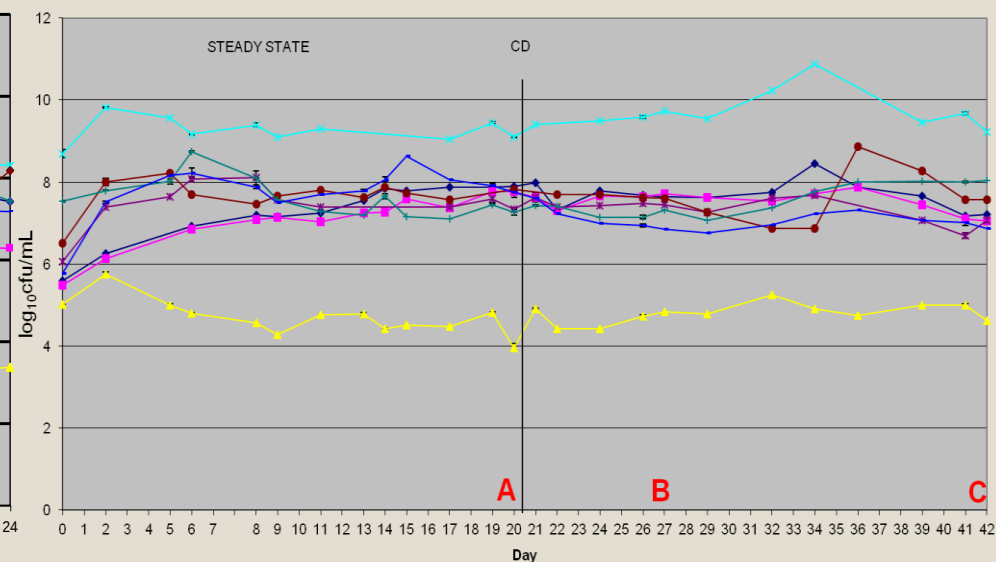
- ❑ Six rods were sampled at three different time points (A, B and C)
- ❑ Planktonic populations monitored throughout
- ❑ Experimental design repeated on separate occasion

Triple stage biofilm human gut model

Planktonic populations of indigenous gut microbiota in vessel 3 of the **original** human gut model

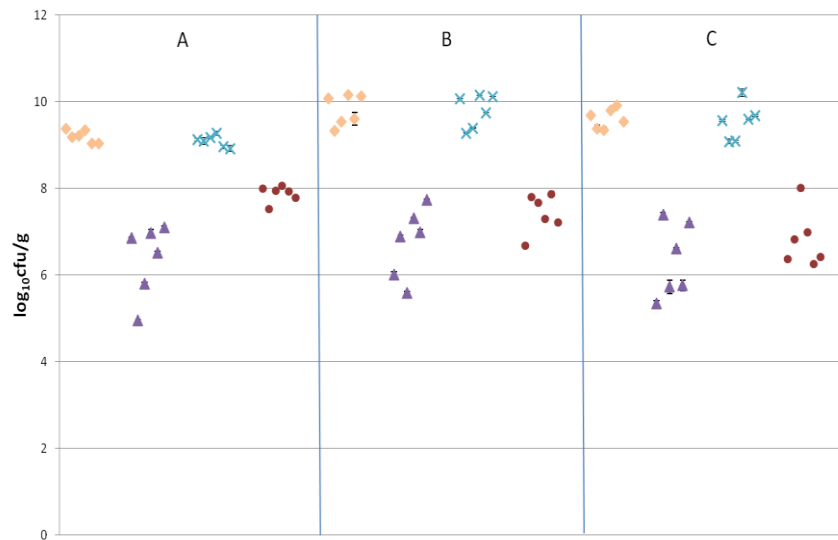


Planktonic populations of indigenous gut microbiota in vessel 3 of the **biofilm** human gut model

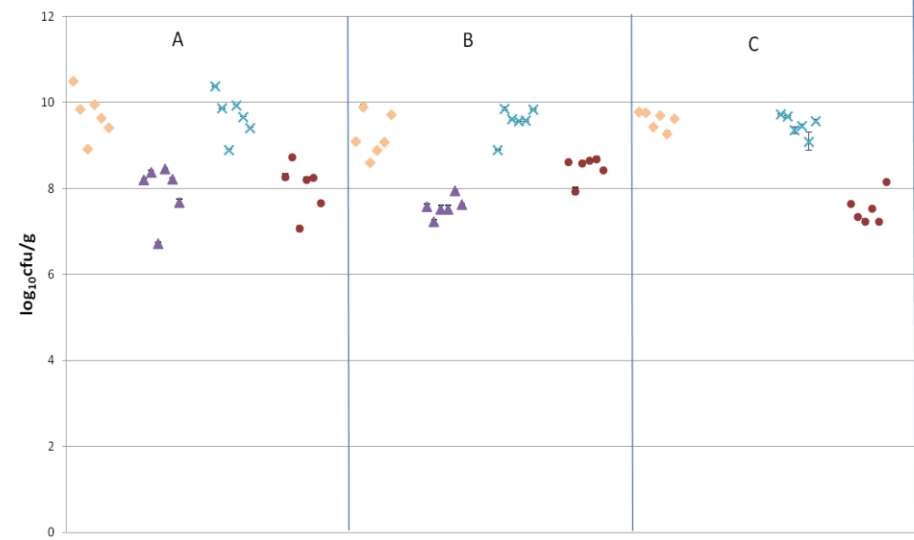


Triple stage biofilm human gut model

Sessile populations of facultative anaerobes within the triple stage biofilm gut model # 1



Sessile populations of facultative anaerobes within the triple stage biofilm gut model # 2



◆ Facultative anaerobes

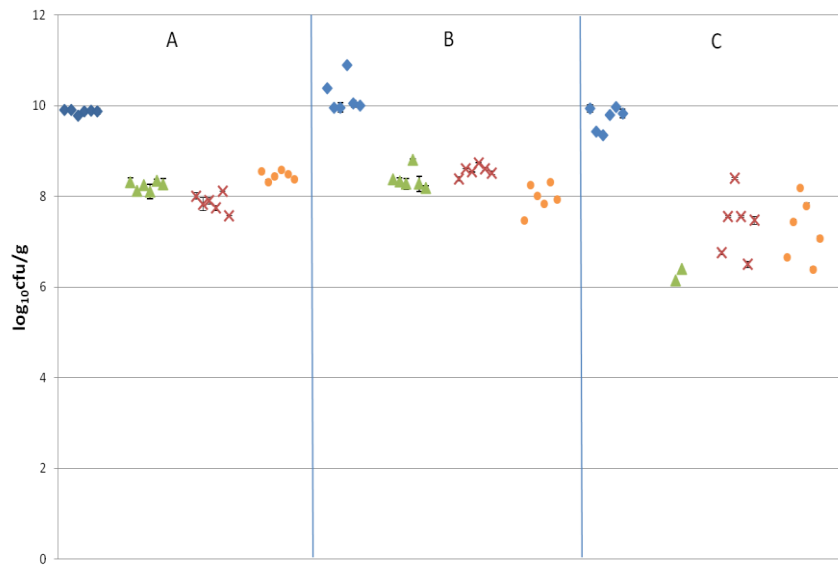
▲ Enterococcus spp

× Lactose-fermenting Enterobacteriaceae

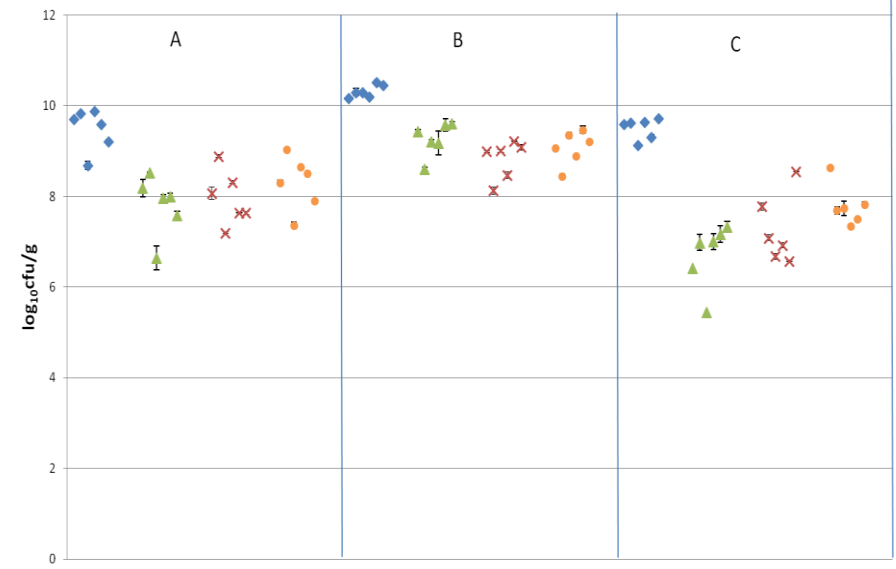
● Lactobacillus spp

Triple stage biofilm human gut model

Sessile populations of obligate anaerobes within the triple stage biofilm gut model # 1



Sessile populations of obligate anaerobes within the triple stage biofilm gut model # 2



◆ Total anaerobes

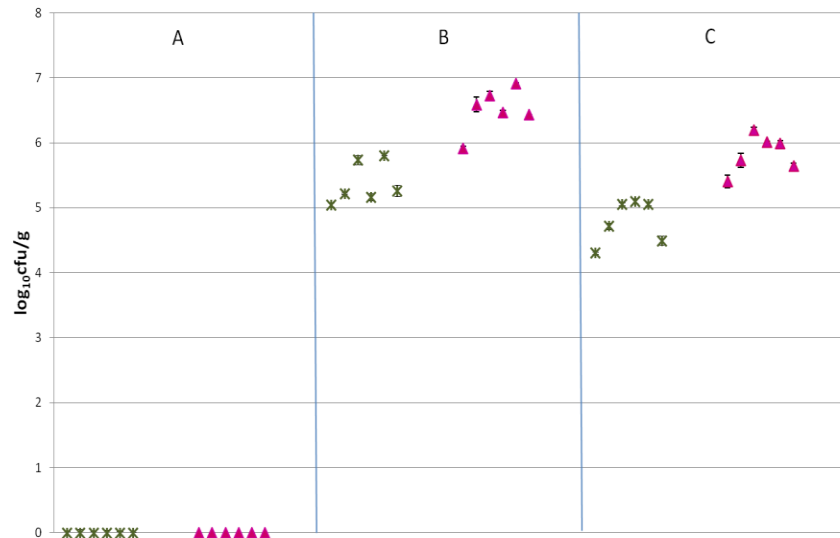
▲ Clostridium spp

× Bacteroides spp

● Bifidobacterium spp

Triple stage biofilm human gut model

Sessile populations of *C. difficile* within the triple stage biofilm gut model # 1

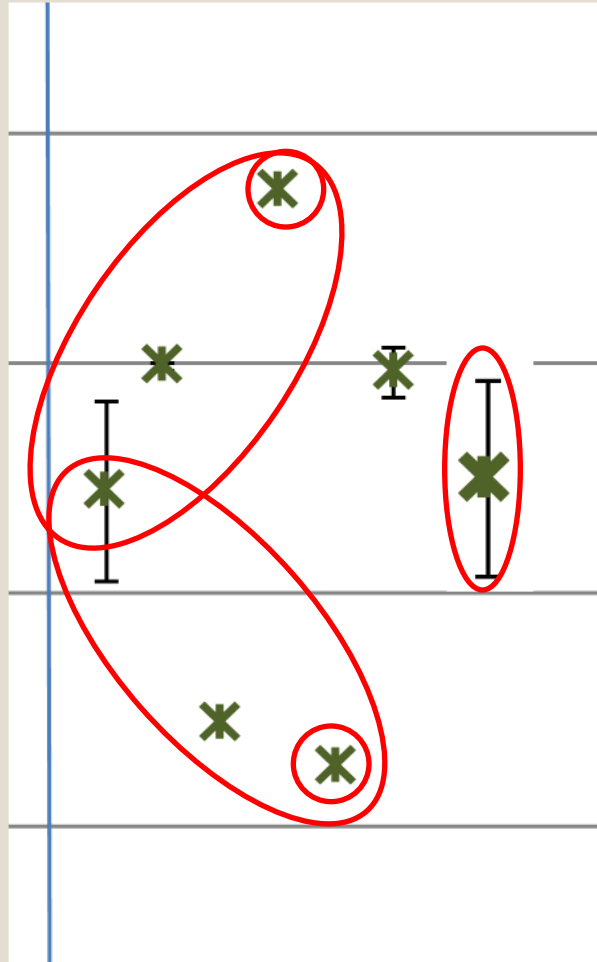


Sessile populations of *C. difficile* within the triple stage biofilm gut model # 2



x C. diff TVC ▲ C. diff spo

Triple stage biofilm human gut model



Triple stage biofilm human gut model

Conclusions

- ❑ Planktonic populations similar to original gut model
- ❑ All bacterial groups present within biofilm including *C. difficile* spores
- ❑ Biofilm forms consistently on all rods
- ❑ Total biofilm biomass is variable
- ❑ Biofilm formation has minimal variation between rods
- ❑ The biofilm model is reproducible
- ❑ Limitation – finite sampling points

Triple stage biofilm gut model has been successfully validated for the study of indigenous gut microbiota biofilms

Future

- ❑ Validate the biofilm human gut model for studying the role of biofilms in CDI
 - Induce CDI
 - Treatment of CDI
 - Post treatment
 - **Does *C. difficile* persist within biofilms after treatment?**
Treatment failure/recurrence?
- ❑ Evaluate drugs for their capacity to treat biofilm infections *in vitro*

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